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Energy Procedia 61 (2014) 2805 – 2808

Energy

**Procedia**The 6<sup>th</sup> International Conference on Applied Energy – ICAE2014

# Energy-efficient extraction of fuel from chlorella combined with CO<sub>2</sub> capture

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## Abstract

To combine the abilities of lipids extraction and CO<sub>2</sub> capture by algae + IL system, chlorella hydrolysis integrating CO<sub>2</sub> removal by ILs ([bmim][BF<sub>4</sub>], [bmim]Cl and [amim]Cl) to extract lipids energy-efficiently was demonstrated in this study. The addition of CO<sub>2</sub> to [bmim][BF<sub>4</sub>] can increase the lipids yield from 14.2% to 15.6%. The value of net energy gain increased from 10.4 to 35.9 with the CO<sub>2</sub> addition to [bmim][BF<sub>4</sub>] because of the compensated CO<sub>2</sub> capture energy in the algae extraction process.

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Peer-review under responsibility of the Organizing Committee of ICAE2014

**Keywords:** fuel extraction; chlorella; CO<sub>2</sub> capture; ionic liquid

## 1. Introduction

Because of the high toughness of bacterial polysaccharide (9 kJ kg<sup>-1</sup>), fuel extraction is the bottleneck for the commercialize of algae-sourced feedstocks [1]. Compared with traditional acid/base water solutions and organic solvents for cell hydrolysis, ILs are generally stable, effective, environmentally benign, and better product selectivity [2]. Meanwhile, room-temperature ILs were proposed as a potential candidate for CO<sub>2</sub> capture in the last few years [3]. Therefore, it would be interesting to take advantage of the abilities of IL for algae hydrolysis and CO<sub>2</sub> capture in the process of fuel extraction from algae. The chlorella hydrolysis integrating CO<sub>2</sub> removal by ILs ([bmim][BF<sub>4</sub>], [bmim]Cl and [amim]Cl) was demonstrated in this study. CO<sub>2</sub> solubility in ILs was measured under the operation conditions of CO<sub>2</sub> capture by ILs (101 kPa, 293 K) and CO<sub>2</sub> release / chlorella hydrolysis (101 kPa, 383 K). The chlorella

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cells hydrolyzed by ILs were observed via transmission electron microscope (TEM). Fuels extraction efficiencies using different IL were compared. Furthermore, the net energy gain from chlorella to lipids, considering energy compensated by CO<sub>2</sub> captured, was estimated.

## 2. Experimental

The experimental schematic was illustrated in Figure 1. The CO<sub>2</sub> solubility in ILs was measured by a self-built apparatus [4] under the operation conditions of CO<sub>2</sub> capture by IL (101 kPa, 20°C) and CO<sub>2</sub> release / chlorella hydrolysis (101 kPa, 110°C). The mixture of chlorella lysis by IL, as shown in Fig.1.c, was sampled for observation *via* TEM (JEM-1400, HITACHI). The contents of biodiesel, as shown in in Fig.1.e, were analysed by a gas chromatograph (GC) (9790 III, FuLi Analytical Instrument). Methyl undecanoate was used as internal standard substance. Extract mass spectra was obtained by ionization time-of-flight (MALDI-TOF) techniques.

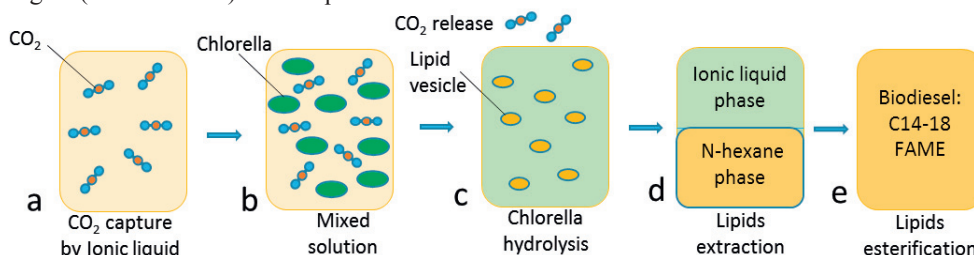


Fig. 1. Experimental schematic.

## 3. Results and discussion

### 3.1. Chlorella experiments

At a volume ratio of 1:10 (wet chlorella : IL), a species comprising a polysaccharide cell wall (cellulosic algae), was hydrolysed by ILs. The lysate of chlorella + [amim]Cl was observed by TEM. Micrographs of intact chlorella and cells broken by hydrolysis of IL were demonstrated in Fig. 2, respectively. The elaborate structure of intact chlorella such as cell wall (Cw), vacuole (V), nucleus (N) and liposome (Ls) can be observed clearly in Fig. 2. (a). Thickness of the intact cell wall was measured to be 109 nm as illustrated in Figure 2 (b). When wet chlorella was mixed with [amim]Cl, part of the cells were disrupted and cytoplasm flowed out from the cells, as shown in Fig. 2 (c). Thickness of the broken cell wall was 47.9 nm illustrated in Fig. 2. (d). Therefore, IL such as [amim]Cl can hydrolysis chlorella cells effectively.

### 3.2. Lipid yield

CO<sub>2</sub> solubility in ILs was presented in Table 1. Under the pressure of 101 kPa, CO<sub>2</sub> was absorbed at the low temperature of 293 K and released at the high chlorella hydrolysis temperature of 383 K. Among three ILs, [bmim][BF<sub>4</sub>] showed the biggest CO<sub>2</sub> solubility of CO<sub>2</sub> with 3.56 mol% at 293 K and 2.78 mol% at 383 K. Yields of lipids from wet chlorella extracted by ILs and accordingly yields of biodiesel were demonstrated in Table 1. The addition of CO<sub>2</sub> to [bmim][BF<sub>4</sub>] can increase the lipids yield from 14.2% to 15.6%. The lipids yield by Soxhlet extractor is 20.8%, which can be roughly used to indicate the total lipids contents in chlorella cells. The value of 15.6% by [bmim][BF<sub>4</sub>] thus means that the percentage

of extracted lipids is 75%. When CO<sub>2</sub> added to the IL, it reacted with water to form H<sup>+</sup> and HCO<sub>3</sub><sup>-</sup>. They may attack on the glycosidic bond. Therefore, an increase in lipids yield by CO<sub>2</sub> addition can be definitely observed for [bmim][BF<sub>4</sub>].

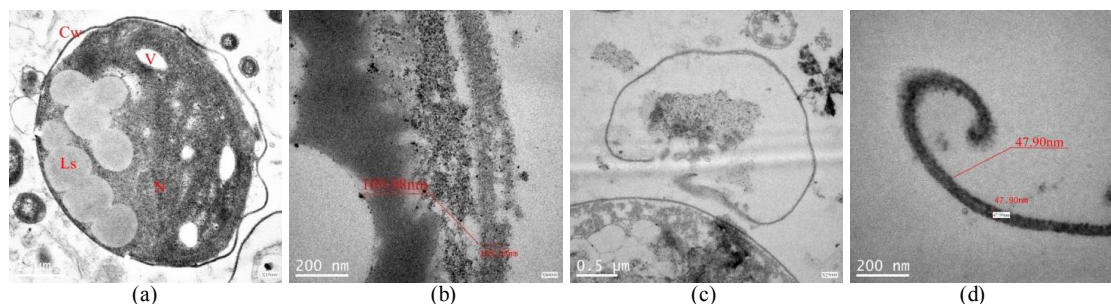


Fig. 2. Ultrastructure of the chlorella cells. (a) intact chlorella cells; (b) wall of the intact chlorella cell (c) chlorella cell hydrolysed by [amim]Cl (d) wall of the chlorella cell hydrolysed by [amim]Cl.

Table 1. CO<sub>2</sub> solubilities, yields of lipids and yields of FAME.

IL	x <sub>a</sub> mol%		x <sub>d</sub> mol%		W <sub>lipid</sub> <sup>a</sup> %		W <sub>FAME</sub> %	
	T=293K	P <sub>CO2</sub> =101 kPa	T=383K	P <sub>CO2</sub> =101 kPa	+ CO <sub>2</sub>	-	+ CO <sub>2</sub>	-
[amim]Cl		2.61		0.23	18.6	18.3	14.2	14.3
[bmim]Cl		3.23		0.25	18.1	17.6	14.2	13.6
[bmim][BF <sub>4</sub> ]		35.6		2.78	15.6	14.2	11.7	10.6

<sup>a</sup>: the lipids yield is calculated by the weight ratio of extracted lipids to dry algae.

### 3.3. Well-to-station net energy gain

The chlorella extraction by [bmim][BF<sub>4</sub>] combining CO<sub>2</sub> capture was employed in the calculation of the net energy gain (NEG) across the “well-to-station” path [2]. Energy consumption and production from chlorella culture to biodiesel combined with energy compensation by CO<sub>2</sub> capture were presented in Table 2. Energy flows for cultivation, harvesting, and transesterification were adopted from the study of Teixeira [2] with wet extraction from nitrogen-starved (Low N). Energy required by CO<sub>2</sub> capture with conventional MEA scrubbing process was calculated to be 3.9 GJ per ton captured CO<sub>2</sub> [5]. The biogas was employed as the feed gas for the calculation of the amount of captured CO<sub>2</sub> [6]. The feed gas was compressed to be 1.2 MPa, and the partial pressure of CO<sub>2</sub> was 0.48 MPa. The electricity energy required by feed gas compression was 0.043 MJ per kg dry algae for work. The corresponding heat energy was -0.044 MJ per kg dry algae for cooling. Those values of 0.043 MJ and -0.044 MJ are much lower than energy compensation by CO<sub>2</sub> captured (1.56 MJ, see Table 2). As shown in Table 2, net energy gain of IL with CO<sub>2</sub> was much higher than that of other campaigns. It should be noted that heat energy cannot be directly compared with electricity energy due to different exergy.

## 4. Conclusion

The addition of CO<sub>2</sub> to [bmim][BF<sub>4</sub>] can increase the lipids yield from 14.2% to 15.6%. The value of net energy gain increased from 10.4 to 35.9 with the CO<sub>2</sub> addition to [bmim][BF<sub>4</sub>] because of the compensated CO<sub>2</sub> capture energy in the algae extraction process.

Table 2. Well-to-station net energy gain (all of values are calculated based on 1kg dry algae).

Process		Conventional extraction		Algae hydrolysis	
		Dry	Wet	Normal	+ CO <sub>2</sub>
Algae culture and harvesting	Electricity (MJ)	1.5	1.5	1.5	1.5
	Heat (MJ)	14	-	-	-
Drying	Electricity (MJ)	1.4	-	-	-
	Heat (MJ)	1.2	2.7-6.6	0.4	0.4
Lipid extraction	Electricity (MJ)	0.3	1.0-2.4	0.1	0.1
	Heat compensation (MJ)	-	-	-	1.56
CO <sub>2</sub> desorption	Heat (MJ)	0.3	0.2	0.2	0.2
	Energy Consumption (MJ)	18	5.4-10.7	2.2	0.64
Lipid transesterification	Energy Production (MJ)	23	23	23	23
	Net energy gain	1.28	2.15-4.26	10.4	35.9

## Acknowledgements

This study was financially supported by the China Natural Science Foundation (Contract No. 21176069), Program for New Century Excellent Talents in University (NCET-10-0380) and the Fundamental Research Funds for the Central Universities (WG1213011).

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## Biography

Junchao Huang is a graduate student in East China University of Science and Technology (ECUST), under the supervision of Prof. Shan-Tung Tu. Junchao Huang's research interests are energy efficient extraction of lipids from algae.